



(19) Europäisches Patentamt
European Patent Office
Office européen des brevets



(11) Publication number: 0 586 183 A1

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 93306733.2

(51) Int. Cl.⁵: G01N 33/58, G01N 33/577,
G01N 33/483, G01N 15/14,
C12Q 1/00

(22) Date of filing: 25.08.93

(30) Priority: 04.09.92 US 940600

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(43) Date of publication of application:
09.03.94 Bulletin 94/10

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(84) Designated Contracting States:
DE FR GB IT

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(54) Control particles for cell counting and instrument linearity.

(57) This invention describes methods and kits for determining instrument linearity of a flow cytometer and is particularly useful as a control particle for use in conjunction with absolute cell counting methods. The particle used in the practice of this invention comprises a small fluorescent bead.

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Field of the Invention

5 This invention relates to the field of control particles products for flow cytometry, and more particularly relates to a method for providing checks on pipetting precision and instrument linearity. The invention is particularly useful in a method for the absolute counting of cells, such as reticulocytes and leukocytes and more particularly one or more subsets of leukocytes, in a cell sample.

Background of the Invention

10 Flow cytometry comprises a well known methodology for identifying and distinguishing between different cell types in a non-homogeneous sample of cells. The sample may be drawn from a variety of sources such as blood, lymph, urine, or may be derived from suspensions of cells from solid tissues such as brain, kidney or liver. In the flow cytometer, cells are passed substantially one at a time through one or more sensing regions wherein each cell is illuminated by an energy source. The energy source generally comprises means that emits light of a single wavelength such as that provided by a laser (e.g., He/Ne or argon) or a mercury arc lamp with appropriate bandpass filters. Different sensing regions can include energy sources that emit light at different wavelengths.

15 In series with each sensing region, various light collection means, such as photomultiplier tubes, are used to gather light that is refracted by each cell (generally referred to as forward light scatter), light that is reflected orthogonal to the direction of the flow of the cells through a sensing region (generally referred to as orthogonal light scatter) and one or more light collection means to collect fluorescent light that may be emitted from the cell as it passes through a sensing region and is illuminated by the energy source. Light scatter is generally correlated with the physical characteristics of each cell.

20 Flow cytometers further comprise data recording and storage means, such as a computer, wherein separate channels record and store the light scatter and fluorescence data from each cell as it passes through a sensing region (i.e., all of the data collected for each cell comprise a "recorded event"). By plotting orthogonal light scatter versus forward light scatter in either real time or by reanalysis of the data after the events have been recorded, one can distinguish between and count, for example, the granulocytes, monocytes and lymphocytes in a population of leukocytes. By gating on only lymphocytes, for example, using light scatter and by the use of appropriate cell markers, such as monoclonal antibodies labelled with fluorochromes of different emission wavelength and/or nucleic acid dyes, one can further distinguish between and count cell types within the lymphocyte population (e.g., between CD4⁺ and CD8⁺ lymphocytes). U.S. Pat. Nos. 4,727,020, 4,704,891 and 4,599,307 describe the arrangement of the various components that comprise a flow cytometer and also the general principles of its use.

25 Because the accurate measurement of light either scattered by or emitted from a cell is critical to the operation of a flow cytometer, assuring that the instrument is properly setup is important to its daily operation and use. There are a number of products available to "setup" a flow cytometer. These products generally comprise polymeric microparticles which are labelled with one or more fluorescent dyes and which come in one or more sizes. The instrument operator has the ability once or more a day to "calibrate" or "align" the instrument 30 using these particles to be sure that the instrument is functioning properly. U.S. Pat. Nos. 5,073,497, 5,073,498 and 5,084,394 describe such beads and methods of use in setting up a flow cytometer. Co-pending and commonly assigned U.S. Serial No. 897,616 (filed June 10, 1992) further describes the use of cells as control particles for staining and instrument calibration.

35 Apart from issues involving the use and operation of the instrument, there are other factors which can effect results gathered by means of flow cytometry. While it is possible using the above-described methods to count the number of cells in a sample and to distinguish between various cell populations, the number of cells counted will be relative (i.e., it will not give an absolute count for a specific volume of blood, for example). Generally, these methods require that red blood cells be substantially removed from the sample. One reason is because the light scatter of the red blood cells and leukocytes is substantially overlapping making their differentiation based on light scatter alone difficult. Another reason is that in order to count leukocytes in a more rapid manner the number of red blood cells must be reduced because the number of red blood cells to leukocytes is approximately 1,000 to 1. Accordingly, practitioners in the field routinely lyse whole blood or separate out the blood cell components by density dependent centrifugation.

40 In addition to the step required for whole blood separation, other steps are routinely involved. For example, once a lysed blood preparation is made, cell markers generally are be added. Unbound markers, then, are routinely washed from the cells. After that step, a fixative is added. Finally, cells in solution are run on a flow cytometer. In some flow cytometers, all of the solution containing cells is delivered to the cytometer for analysis. In other flow cytometers, only a measured amount of solution is delivered.

While lysis and washing steps are routine, there are now methods and procedures that do not involve washing. These so-called "no-wash" methods involve the addition of immunofluorescence markers and fixatives without washing steps. Co-pending and commonly assigned U.S. Serial No. 846,316 (filed March 5, 1992) is directed to one such method.

5 Regardless of the method or instrument used, any step in which the sample of cells is physically manipulated introduces not only the possibility for error, but also the loss of cells from the sample. In addition, each step increases the risk to the technician of being exposed to contaminated blood. Using these traditional flow cytometric methods, therefore, the number of cells in a given volume of blood cannot be easily or accurately determined.

10 Thus, in each of the presently described systems, there are one or more obstacles that prevent one from assuring instrument accuracy and/or accurately determining the absolute count of specific cells in a heterogeneous sample of blood. These obstacles are not overcome by the mere addition of a reference particle, as described in U.S. Pat. No. 4,110,604, with flow cytometry or the occasional calibration of the instrument with control particles. Several drawbacks remain.

15 A major drawback to the use of flow cytometers is that unless the fluorescence channels and optical alignment of each flow cytometer is calibrated to read the same, there is no assurance as to the source of variation in a sample. It is likely that one instrument will give different readings on the same sample on different days if it was aligned and/or calibrated differently each day. Similarly, there is no assurance that any two instruments will provide the same results even if properly set up. Accordingly, while flow cytometry provides a better measure 20 of identifying and distinguishing between cells in a sample, its present use as a clinical instrument may be diminished by the limitations in set up and operation if not properly performed. What is required is a single system or method that will allow one to accurately count cells in a sample and/or be assured that the results from one instrument are consistent from sample to sample as well as consistent with results obtained from other instruments.

25 Even through the resolution of some of these problems through the methods and kits described in co-pending and commonly assigned U.S. Serial No. 570,569 (filed August 7, 1990), additional issues remain. For example, the use of a single reference particle will provide a means to perform absolute counts in a cell sample. The addition of such particles, however, does not assure that the technician handling the samples or the equipment used to handle the samples are accurate. Sample error introduced through the pipetting step can be a 30 source of significant variation which cannot be otherwise accounted for. Similarly, the use of a single reference particle may not assure that the flow cytometer is accurately counting the particles, and thus, the cells. If the particle count is not accurate, the cell count cannot be accurate.

The present invention overcomes all of these obstacles and provides a one step test for absolute counting of one or more specific populations of cells in an unlysed whole blood sample. The present invention further 35 provides a check on pipetting accuracy and provides a check on instrument linearity.

Summary of the Invention

40 This invention, therefore, relates to a method and kit comprising control reagents for conventional (i.e., non-absolute) cell counting. In this method, a test sample is split into more than one aliquot, preferably two, more preferably four, and each aliquot is added to a separate tube. Each tube may contain one or more cell markers or the markers may be added as a separate step. To each tube is added a known amount of microparticles. The amount of microparticles in each tube will differ. Each tube then is analyzed by means of flow 45 cytometry and the number of microparticles and cells per tube is counted. The result from counting should be linear if pipetting accuracy and instrument linearity are correct. In the instance where two or more cell markers are being used to identify two or more subsets of cells, it is preferable to use at least four tubes and to add one cell marker to two tubes and the other cell marker to the two other tubes. In this instance, if the instrument is functioning properly, the coefficients of variation between tubes containing the same marker should be minimal.

50 The invention has particular utility in a method and kit for the absolute counting of one or more populations of cells in a sample. The preferred means for counting such cells comprises a flow cytometer. In this embodiment of the method, a test sample is added to a tube. The tube may contain a diluent. The diluent may comprise a mixture of a fixative, one or more cell markers and a known amount of a first microparticle. The first microparticle is fluorescent and the fluorescence is distinguishable from the fluorescence emitted by the cell marker(s). To this mixture, a known amount of a second fluorescent microparticle is added. The fluorescence of 55 the second microparticle is distinguishable from the fluorescence emitted by the cell marker(s) and the first microparticle. The sample then is vortexed, incubated, vortexed again and run on a flow cytometer having one or more fluorescence channels.

In an alternative embodiment, the diluent may comprise only the cell markers. In that instance, the fixative and first microparticles are added separately. In another embodiment, the diluent may comprise the cell markers and first microparticles. The fixative is added separately. The timing of the addition of these various components is not critical to the practice of the invention.

5 Fluorescence data is recorded and stored for each event. A fluorescence trigger is set for one fluorescence channel so as to include essentially all of the microparticles and cells to be counted. The number of microparticles then is counted by analyzing the recorded events.

10 Counting the number of cells in the sample, the number of first microparticles and by knowing the amount of first microparticles added per unit volume, the number of cells in each population can be absolutely counted. Counting the number of second microparticles and the number of first microparticles in the sample permits a check on pipetting accuracy and instrument linearity.

15 A kit useful in the practice of this invention comprises the following items: a sample tube and a diluent wherein the diluent comprises a mixture of one or more cell markers and a known amount of a first microparticle. The kit also will contain two or more containers having different concentrations of a second microparticle. The diluent may be packaged in the tube. In the tube, the diluent may be liquid or may be dried by methods known to those skilled in the art such as lyophilization. Drying may be performed in the presence of a stabilization agent such as trehalose. In the dried format, the "diluent" will be return to solution upon addition of the liquid sample such as blood. In any embodiment, the diluent may be separately contained or may be broken up into its several components each of which may be separately contained. In these alternatives, the diluent 20 may be added to the sample tube before or after the sample is added to the tube.

Another kit useful in the practice of this invention comprises two or more tubes and two or more containers each container having a different amount of a fluorescent microparticle contained therein.

Brief Description of the Drawings

25 FIG. 1 comprises four dot plots of log fluorescence 2 versus log fluorescence 1 for recorded events exceeding a first fluorescence trigger on the log fluorescence 1 axis from unlysed whole blood cells split into four aliquots and stained with a diluent containing in (A) and (B) anti-CD3(PE) and anti-CD4(PE/CY5) and in (C) and (D) anti-CD3(PE) and anti-CD8(PE/CY5). In all four tubes, the diluent contained the same amount of 30 a first fluorescent microparticle. In (A), 0 second microparticles per μ l were added; in (C), 56 second microparticles per μ l were added; in (B), 278 second microparticles per μ l were added; and in (D), 1115 second microparticles per μ l were added.

Detailed Description

35 The invention comprises methods and kits for determining instrument linearity and pipetting accuracy and is particularly useful as part of a method for determining the absolute count for one or more populations of cells in a sample, preferably, by flow cytometry. In either instance, the sample may be derived from any tissue source or cell line, but generally is selected from the group consisting of whole blood, lymph, spinal fluid, urine and bone marrow. Unlysed whole blood is preferred as a source of cells for determining instrument linearity.

40 The populations of cells that can be counted in a sample comprise platelets, red blood cells, white blood cells and subsets and precursors of each. One preferred population of red blood cells comprise reticulocytes. The preferred subsets of leukocytes comprise lymphocytes, monocytes and granulocytes. In one preferred embodiment, lymphocyte subsets are particularly important and more preferred is the counting of CD4 $^{+}$ /CD3 $^{+}$ and CD8 $^{+}$ /CD3 $^{+}$ T lymphocytes in an unlysed whole blood sample. It will be appreciated that this invention is applicable to counting any population of cells (e.g., CD34 $^{+}$ stem cells) as well as to any number of populations of cells. For example, in order to count both the number of CD4 $^{+}$ and CD8 $^{+}$ T lymphocytes in a sample, one would use anti-CD4 and anti-CD8 antibodies. In such an instance, it is preferable to use anti-CD3 antibodies as well 45 count only those CD4 and CD8 cells that are lymphocytes. In another example, to calculate a three part white blood cell differential, one could use anti-CD45, anti-CD14 and anti-CD15 antibodies. In still another example, to calculate the absolute number of T lymphocytes and/or B lymphocytes, one could use anti-CD3 and/or anti-CD19 (or anti-CD20) antibodies to count T cells or B cells respectively. Any population that can be identified 50 by a single cell marker can be counted alone or can be counted with other populations in the same sample.

55 Cell markers useful in the practice of this invention comprise immunofluorescence markers and other fluorescence tagging agents which will specifically label one or more populations of cells. As set forth above, immunofluorescence markers comprise antibodies bound to fluorochromes. Monoclonal antibodies are preferred. Examples of fluorescence tagging agents include nucleic acid dyes, such as those described in U.S. Pat No.s 4,544,546, 4,883,867 and 4,937,198, and such dyes as propidium iodide, acridine orange, thiazole or-

ange, thioflavin T and 7-amino-actinomycin D. A preferred nucleic acid dye described generally by formula I of U.S. Pat. No. 4,544,546 having a quinoline nucleus is presently marketed as a laser dye under the tradename LDS-751 (Exciton).

5 Fluorochromes useful in the practice of this invention may be excitable at the same wavelength of light. Dyes having these properties include the phycobiliproteins (especially phycoerythrin "PE"), fluorescein derivatives (such as fluorescein isothiocyanate "FITC"), peridinin chlorophyll protein complex (such as described in U.S. Pat. No. 4,876,190 "PerCp"), coumarin derivatives (such as aminomethyl coumarin), pthalocyanine dyes (such as Ultralite dyes (Ultradiagnostics)), rhodamine derivatives (such as tetramethyl rhodamine or Texas Red (Molecular Probes)) and cytofluorochromes (such as CY3 and CY5 (Biological Detection Systems)).

10 Tandem conjugates formed between two of the dyes set forth above also may be useful within the practice of this invention. See U.S. Pat. No. 4,542,104.

15 When more than one population of cells is to be counted, more than one cell marker may be used (each being specific for a different population); however, the fluorescence of each marker must have emission wavelengths that are distinguishable not only from each other but also from the microparticles used. When only one immunofluorescence marker is used, PE is preferred as the fluorochrome. When two or more immunofluorescence markers are used, PE and a tandem conjugate formed between PE and CY5 is preferred.

20 The microparticle used in the practice of this invention should have certain properties. First, it should be small (i.e., between 0.2 μ m and 20 μ m, with 4 μ m preferred) so as to stay suspended in the mixture and not settle any faster than the cells in the sample. Second, it should be made of a material that avoids clumping or aggregation. Polystyrene is preferred. Third, it should be fluorescent. Fluorescence can be achieved by selecting the material that comprises the microparticle to be autofluorescent or it can be made fluorescent by being tagged with one or more fluorescent dyes to appear autofluorescent. A combination of red and blue emitting dyes are preferred.

25 The fluorescence of the microparticles must be such that it is sufficiently greater than noise from background so as to be distinguishable. When used with one or more cell markers and/or with another microparticle (i.e., first microparticle), it also must be distinguishable from the fluorescence emitted by such marker(s), and/or from the fluorescence emitted by another microparticle. Three standard deviation difference between that fluorescence and the microparticle fluorescence is sufficient. Microparticles having these properties may be selected from the group consisting of fixed chicken red blood cells, coumarin beads, liposomes containing a fluorescent dye, fluorescein beads, rhodamine beads, fixed fluorescent cells, fluorescent cell nuclei, micro-organisms and other beads tagged with a fluorescent dye. Polystyrene microparticles tagged with red and blue emitting dyes are preferred.

30 The concentration range of the microparticle should be sufficient to demonstrate linearity and pipetting accuracy. The lower endpoint of the range is preferably 0 particles per unit volume while the upper endpoint of the range should be at or near the concentration of cells in a sample to be counted. Generally, two different concentrations of particles is preferred; three different concentrations is more preferred; and four different concentrations is most preferred. When more than two concentrations are used, it is preferred that at least two of concentrations be at or near the endpoints.

35 In one embodiment of the invention, therefore, identical volumes of each of the different concentrations of microparticles are added to different tubes containing equal volumes of a sample. Thus, in the most preferred embodiment, one tube will contain 0 particles per μ l, one tube will contain a concentration of particles at or near the number of cells to be counted in a sample (e.g., 1000 per μ l), and each of the two remaining tubes will contain different concentrations in between (e.g., 55 particles per μ l and 275 particles per μ l).

40 The flow cytometer should be equipped with one or more fluorescence detectors (arbitrarily referred to as fluorescence channels 1 and 2 or "FL1" and "FL2" etc.) and with data recording and analysis means, such means generally comprising a computer. Each of the tubes then are run through the flow cytometer. Fluorescence and scatter data for each event is recorded. A fluorescence trigger is set such that essentially all the microparticles to be counted meet or exceed the trigger level. In the preferred embodiment, the trigger is set to include at least 99%, and preferably 99.99%, of all microparticles to be counted. The data is recorded and an analysis of the data is made to determine if the instrument is counting the particles in the different tubes in a linear manner and if pipetting is accurate. The data may be analyzed by simple statistical analysis such as standard regression analysis.

45 In another embodiment of the invention, the particles are added to two or more aliquots of a test sample containing cells. Where only a single cell marker is being used, two different concentrations of the particles may be used although, again, four is preferred for statistical reasons. Where two or more cell markers are being used, it is preferable to use at least four different concentrations of particles.

50 The following example describes each embodiment of the invention in more detail and makes use of the methods described in U.S. Serial No. 570,569.

Unlysed whole blood was divided into ten replicates of four aliquots each. In the four aliquots, two ((A) and (B)) contained a diluent comprising anti-CD3(PE/CY5) and anti-CD4(PE) and the other two tubes ((C) and(D)) contained anti-CD3(PE/CY5) and anti-CD8(FE). All four tubes contained a first microparticle consisting of a polystyrene microsphere containing orange emitting dyes at a concentration of 150 per μ l.

5 A second microparticle consisting of a polystyrene bead containing red and blues dyes was set used to establish a range of concentrations. The concentrations selected were 0, 56, 278 and 1115 particles per μ l. 50 μ l of a concentration was added to each of the four aliquots: 0 to (A); 278 to (B); 56 to (C); and 1115 to (D). The tubes were vortexed, a fixative was added and each tube was analyzed on a FACScan brand flow cytometer (BDIS). The instrument was equipped with LYSIS II software (BDIS).

10 Referring to FIG. 1, the results of the analysis are shown. In (A), the area circled and labelled "R3" contains those cells that are CD3 $^{+}$ but not CD4 $^{+}$. "R2" contains those cells that are CD3 $^{+}$ and CD4 $^{+}$. "R1" contains the first microparticles and "R4" contains the second microparticles. The unlabelled circles in (B), (C) and (D) have same meanings as in (A) except that CD4 is replaced by CD8 in (C) and (D).

15 Comparing (A) with (B), it is apparent that the only visual difference in these plots is in the contents of the circle labelled R4 because the number of CD4 and CD3 cells should not vary between the tubes nor should the number of first microparticles. The same is true when (C) and (D) are compared. When all four aliquots are compared, there are no visual differences between each of the R1 areas nor between each of the R3 areas because each should contain the same number of CD3 cells and first microparticles if the instrument is properly aligned and pipetting of the reagents was accurate. Table I summarizes the actual counts for CD4/CD8, CD3 and second microparticle from each replicate.

TABLE I

25	TUBE	REP#	CD4/CD8 COUNT	CD3 COUNT	2ND PARTICLE COUNT	2ND PARTICLE INPUT
30	A	1	700	999	0	0
	A	2	696	1011	0	0
	A	3	670	968	3	0
	A	4	700	1007	1	0
	A	5	668	966	0	0
	A	6	663	980	1	0
	A	7	684	969	1	0
	A	8	684	998	1	0
	A	9	676	977	1	0
	A	10	691	992	0	0
45	B	1	674	960	266	278
	B	2	689	990	287	278
	B	3	667	957	271	278
	B	4	694	985	274	278
	B	5	685	984	288	278
	B	6	644	918	282	278
	B	7	662	963	291	278
	B	8	659	951	269	278

TABLE I cont.

5	B	9	667	960	264	278
	B	10	698	1011	280	278
10	C	1	245	989	62	56
	C	2	239	958	59	56
	C	3	252	953	52	56
	C	4	241	922	54	56
15	C	5	255	1015	57	56
	C	6	234	943	54	56
	C	7	255	986	62	56
20	C	8	252	1006	55	56
	C	9	257	1014	68	56
	C	10	248	942	60	56
25	D	1	248	967	1175	1115
	D	2	248	959	1101	1115
	D	3	251	993	1087	1115
30	D	4	241	932	1085	1115
	D	5	254	996	1181	1115
	D	6	229	908	1073	1115
35	D	7	247	945	1114	1115
	D	8	234	916	1108	1115
	D	9	250	951	1114	1115
	D	10	238	947	1114	1115

40 Taking the data tabulated in Table I, the mean, standard deviation ("S.D") and coefficient of variation ("CV%") were calculated for CD4/CD8, CD3 and second microparticle in each of tubes (A), (B), (C) and (D), for all CD4 cells (i.e., (A) and (B)), for all CD8 cells (i.e., (C) and (D)), and for all CD3 cells (i.e., (A), (B), (C) and (D)). The results are summarized in Table II.

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TABLE II

TUBE		CD4/CD8	CD3	2ND PARTICLE
5	A	Mean	683	987
		S.D.	13	16
		CV%	1.9	1.6
10	B	Mean	674	967
		S.D.	16	24
		CV%	2.4	2.5
15	C	Mean	247	973
		S.D.	8	32
		CV%	3.1	3.3
20	D	Mean	244	952
		S.D.	8	28
		CV%	3.2	2.9
25	CD4	Mean	679	977
		S.D.	15	23
		CV%	2.3	2.3
30	CD8	Mean	245	962
		S.D.	8	32
		CV%	3.2	3.3
35	CD3	Mean	970	N.A.
		S.D.	28	
		CV%	2.9	

50 As expected, CD4 and CD8 absolute counts are consistent between replicates within a tube. The coefficient of variation of all forty total CD3 determinations was 2.9%. By having four determinations made, the CD3 count can be used to detect pipetting inaccuracies.

55 For the linearity determinations, linear regression analysis yielded an R^2 value of 0.998 with a slope of 0.999 ± 0.006 . Thus, any R^2 value which differed significantly from 1 would detect a deviation from linearity. Similarly, pipetting accuracy would be reflected in an altered regression slope.

All publications and patent applications mentioned in this specification are indicative of the level of ordinary skill in the art to which this invention pertains. All publications and patent applications are herein incorporated

by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

It will be apparent to one of ordinary skill in the art that many changes and modifications can be made in the invention without departing from the spirit or scope of the appended claims.

5

Claims

1. A method for determining instrument linearity of a flow cytometer comprising the steps of:
 - 10 (a) separating a sample of cells into at least two aliquots;
 - (b) adding an equal volume of fluorescent microparticles to each of the aliquots wherein the concentrations added to each aliquot differ;
 - (c) counting the number of microparticles added by means of flow cytometry; and
 - (d) performing statistical analysis on the results to determine linearity.
- 15 2. In a method for absolute counting of cells in a sample wherein the sample is mixed with one or more cell markers and a first fluorescent microparticle, a method comprising the steps of:
 - (a) separating the sample of cells into at least two aliquots;
 - (b) adding an equal volume of a second fluorescent microparticles to each of the aliquots wherein the concentrations added to each aliquot differ;
 - 20 (c) counting the number of fluorescent cells, the number of first microparticles and second microparticles by means of flow cytometry; and
 - (d) performing statistical analysis on the number of cells counted, the number of first microparticles counted and the number of second microparticles counted to determine linearity.
- 25 3. The method of claim 2 wherein the second fluorescent microparticle is selected such that the fluorescence emitted from the second microparticle is distinguishable from the fluorescence emitted from the first microparticle and cell markers.
- 30 4. The method of claim 2 wherein the cell markers comprise fluorescently labelled monoclonal antibodies.
5. The method of claim 4 wherein the cell markers comprise fluorescently labelled anti-CD4, anti-CD8 and anti-CD3 monoclonal antibodies.
- 35 6. The method of claim 4 wherein the fluorescent dyes are selected from the group consisting of phycobiliproteins, fluorescein derivatives, peridinin chlorophyll protein complex, coumarin derivatives, pthalocyanine dyes, rhodamine derivatives, cytofluorochromes and tandem conjugates formed between two of the dyes set forth herein.

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FIG-1A

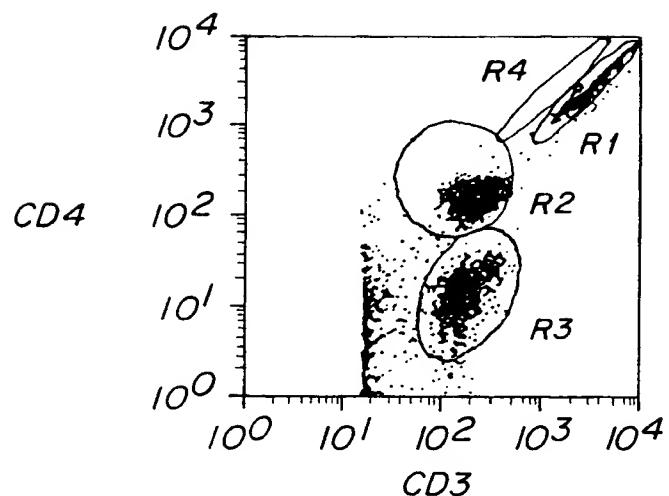


FIG-1B

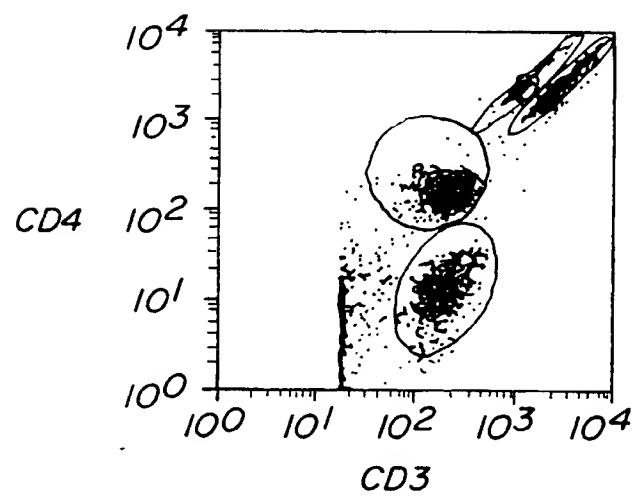


FIG-1 C

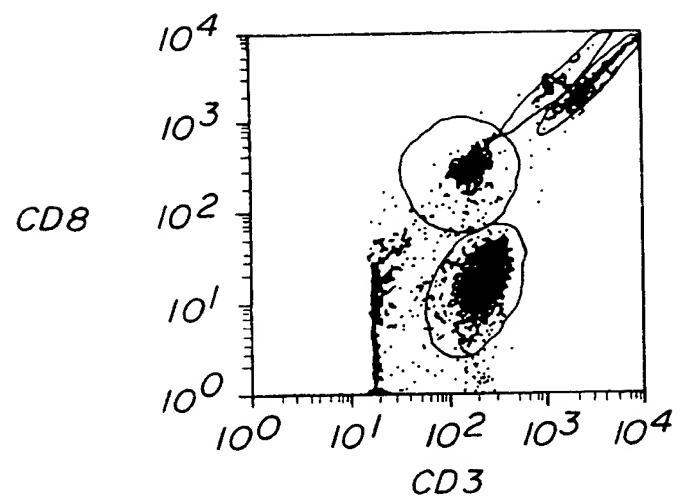
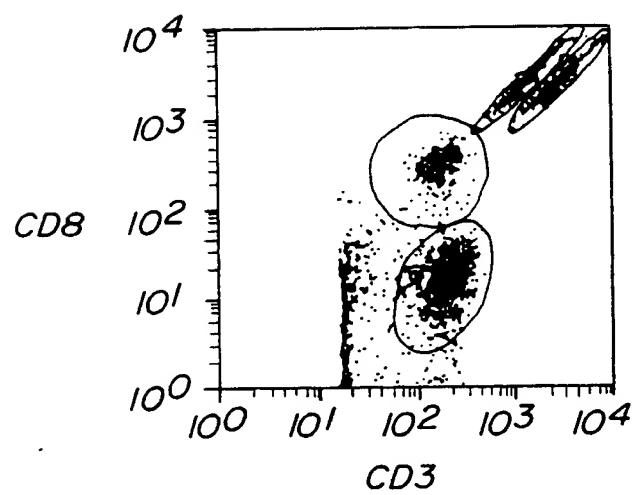


FIG-1 D





DOCUMENTS CONSIDERED TO BE RELEVANT			EP 93306733.2
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
D, A	<u>EP - A - 0 470 810</u> (BECTON, DICKINSON & COMPANY) * Claims * --	1-6	G 01 N 33/58 G 01 N 33/577 G 01 N 33/483 G 01 N 15/14 C 12 Q 1/00
D, E, A	<u>EP - A - 0 559 208</u> (BECTON, DICKINSON & COMPANY) * Claims * --	1-6	
D, A	<u>US - A - 5 084 394</u> (VOGTT et al.) * Claims * --	1	
D, A	<u>US - A - 5 073 497</u> (SCHWARTZ) * Claims * -----	1	
TECHNICAL FIELDS SEARCHED (Int. Cl.5)			
G 01 N C 12 Q			
The present search report has been drawn up for all claims			
Place of search VIENNA	Date of completion of the search 17-11-1993	Examiner SCHNASS	
CATEGORY OF CITED DOCUMENTS			
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document	